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Efficacy of Paromomycin Sulphate for the Treatment of Cryptosporidiosis in Goat Kids

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ABSTRACT

Background: This study aimed to evaluate the therapeutic efficacy of paromomycin sulphate on serum amyloid A (SAA) and haptoglobin (Hp) in Cryptosporidiosis in goat kids.

Methods: In the study, stool samples were taken from the kids and Cryptosporidium was detected via the rapid test kit. Then, by examining the same stool samples microscopically, we included 9 kids in the study and 50 mg/kg of paromomycin sulphate was administered to the kids in which *Cryptosporidium spp.* was detected for five days as a treatment. For the SAA and Hp estimation, blood samples were drawn from the kids in the pre and post treatment stages.

Result: In the kids with Cryptosporidium, the concentrations of SAA and Hp were measured as (SAA; 17.10±4.18 ng/mL Hp; 2.68±1.08 µg/mL) in pre-treatment and as (SAA; 9,71±4,13 ng/mL, Hp; 1,23±0,64 µg/mL) in the post-treatment stage ($p<0,05$). In accordance with these findings, we suggest that routine measurements of SAA and Hp concentrations of the goat kids with diarrhea due to cryptosporidiosis could be of value in follow up of treatment, determining the severity of the infection, selection of therapy and monitoring the efficacy of the preferred treatment protocol and in detection of subclinical infections.

Key words: Cryptosporidiosis, Goat kids, Haptoglobin, Serum amyloid A.

INTRODUCTION

Neonatal Diarrhea Syndrome is a common issue in sheep and goat herds and Cryptosporidiosis, which also contributes to that syndrome, has been reported to be the one of the reasons of the enteric disease in humans and farm animals (Kabu *et al*., 2023; Feng and Xiao, 2017; Giadinis *et al.,* 2012). Among the *Crytospordium* species, *Cryptosporidium parvum, C. ubiquitum* and *C. xiaoiare* are the dominant species in the sheep and goats. *C. parvum* is one of the most pathogenic species while, *C. ubiquitum* is less common in regions where *C. parvum* is endemic. *C. xiaoi* has a narrower host range owing to its non-pathogenic character and nature (Guo *et al.,* 2021). Besides this, some of *Crytosporidium* isolates are zoonotic (Giadinis *et al.,* 2015; Cacciò *et al.,* 2013). Due to the high mortality and morbidity rate associated with its infection, this protozoan parasite has been reported to cause economic losses (Giadinis *et al.,* 2015). It has also been reported that *Crytosporidium* transmits both directly and indirectly *via* fecal-oral path; direct transmission occurs when fecal oocysts are swallowed whereas indirect transmission involves the consumption of oocysts contaminated food and water (Santin, 2020). It has been stated that *Crytosporidium spp*. has a significant impact on the health of 4-15 days old kids; although the animals have a normal appetite at the beginning of the infection, they may die after developing diarrhea, dehydration and electrolyte imbalance (Giadinis *et al.,* 2015; Giadinis *et al.,* 2012). Moreover, it has been asserted that when other enteropathogens become more complicated with *Cryptosporidium* infection, the course of infection gets shorter, ending in death within 2 to 3 days (Wright and Coop, 2007).

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Acute phase response (APR) is defined as a non-specific and complex reaction which occurs shortly after the tissue damage caused by various conditions such as infection, trauma, neoplasia, inflammation and stress (Criscitiello *et al.,* 2019). APR has been reported to include countless symptoms like edema, erythema and fever, which develop due to a complex interaction between cytokines, chemokines and acute phase proteins (APPs), leukocytosis, increased secretion of various hormones along with the regulation of plasma protein synthesis (Criscitiello *et al.,* 2019; Cray, 2012). It has been ascertained that although APPs are mainly synthesized by hepatocytes that are stimulated by cytokines or endogenous glucocorticoids, they can be produced by other tissues as well (Criscitiello *et al.,* 2019; Ceciliani *et al.,* 2012; Cray, 2012). In response to bacterial and parasitic infections, the host may also cause acute phase response, which causes many systemic effects

such as stimulation of fever, cachexies and acute phase proteins (Ngure *et al*., 1997). In ruminants, HP and SAA are seen as the main source of acute phase response in a positive direction (Lecchi *et al*., 2012). It has been reported that SAA, which is a major APP, has three distinct aspects including the binding of cholesterol, immunomodulator activity and opsonization while Hp is an indicator of chaperone activity owing to binding free hemoglobin, anti-inflammatory characteristic, bacteriostatic impact and its role in angiogenesis (Cray, 2012). It has been reported that SAA and Hp concentrations in goats increased in helminth infections, pregnancy toxemia, subacute ruminal acidosis and experimental mastitis (Heller and Johns, 2015; Cray, 2012).

The purpose of this study was to determine the changes caused by cryptosporidiosis in SAA and Hp concentrations in neonatal goat kids after treatment with paramomysin sulphate and to evaluate the therapeutic efficacy of the treatment.

MATERIALS AND METHODS

In order to determine the changes in SAA and Hp values in goat kids in the pre and post treatment stages, nine 3-15 days old goat kids with clinical diarrhea were included in the present study. In fresh stool samples taken from diarrhea kids, *Cryptosporidium* spp. was detected by using antigen-sensitive fast test kits (Rota-Corona-*E. coli*- Cryptosporidium) (Bio-X Diagnostics S.A. RoheFort/Belgium).

The stool samples in which Cryptosporidium spp. was detected by using test kits in the field were transported to the laboratory within the same day. To detect the oocysts of Cryptosporidium spp., stool samples were stained by using Kinyoun Acid Fast staining method (Turgay, 2011). Stained samples were examined under Olympus CX31 trinocular research microscope at magnifications of 10x and 40x. The oocysts detected in the samples were photographed by using the Olympus LC_{30} digital camera system (Fig 1).

Blood samples (5 ml) taken from vena jugularis of kids with diarrhea drawn in vacuum with clot activator serum tubes before treatment were were centrifuged at room temperature at 5000 rpm and serum was collected. For treatment of cryptosporidiosis 50 mg/kg of paromomycin sulphate (Parafor, Huvepharma®) was administered orally for five days. After the treatment, the blood samples taken from the jugular veins of the kids into vacuum tubes with clot activator were centrifuged at 5000 rpm at room temperature to obtain serum. Serum samples were stored at -20°C until the measurement. Serum Amyloid A (Cusabio Biontech CO., LTD. China) and Haptoglobulin (Cusabio Biontech CO., LTD. China) were estimated using made in commercial sandwich ELISA kits.

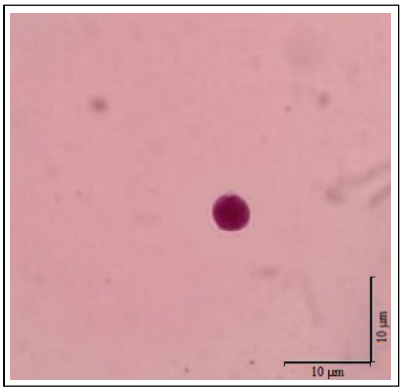


Fig 1: *Cryptosporidium* spp. oocyst in faecal sample stained by using Kinyoun acid fast staining method.

For statistical analysis, the data was analysed with Wilcoxon test and the pre and posttreatment stage SAA and Hp values of the kids were compared. The significance level was set as $p<0.05$.

RESULTS AND DISCUSSION

In our study, SAA concentrations in goat kids with *Cryptosporidium* spp. infection were determined in pre (17.10±4.18 ng/mL) and post (9.71±4.13 ng/mL) treatment stages. In the light of these results, the comparison of SAA measurements of kids revealed a statistically significant difference ($p<0.05$) between pre-treatment values and post-treatment (9.71±4.13 ng/mL). SAA value was much higher in pre-treatment stage than in post-treatment phase (Table 1).

Similarly, the comparison of Hp concentration revealed values as in pre-treatment (2.68±1.08 µg/mL) and in post-treatment (1.23±0.64 µg/mL). According to these findings, there was a statistically significant difference between the pre) and post treatment stages Hp value was higher in pre-treatment than in the post-treatment phase. When the estimated values are considered, it is obvious that the applied treatment apparently and positively affected both SAA and Hp concentrations, which indirectly proves the efficacy of the treatment (Table 1).

Although there are many studies on the values of APPs in Neonatal Diarrhoea Syndrome in calves and lambs, but no literature is available on the level APPs in goat kids affected with Neonatal Diarrhoea Syndrome (Kabu *et al*., 2023; Kabu and Uyarlar, 2022; Dinler *et al.,* 2017; Albayrak and Kabu, 2016). Thus, results of our study Indicate that APPs can be used as a potential diagnostic tool in veterinary medicine. Showing a faster response to an experimental inflammatory stimulus than other APPs in goats, SAA can be a major APP (increase up to 22 folds) indicating an acute condition (González *et al.,* 2008). In a study on goats naturally infected with paratuberculosis, SAA concentration was reported to increase (Sevgisunar and Şahinduran, 2021). Demirbas and Kabu (2021) determined in their study on pneumonic goats that SAA concentration was higher in the pneumonic group than the healthy goats. Kabu *et al*., (2023), SAA and Hp concentrations were found high in lamb with diarrhea due to Cryptosporidium. Additionally, SAA and Hp concentrations were found high in goats with pregnancy toxemia (Albay *et al.,* 2014). In a study on the impacts of temperature and transport stress on SAA and Hp concentrations in goats, SAA and Hp concentrations were

Table 1: Pre and post treatment concentrations of SAA and Hp in goat kids with *Cryptosporidium* spp.

Group	Pre-treatment	Post-treatment	*P*
SAA (ng/mL)	17.10±4.18[a]	9.71±4.13[b]	0.021
Hp (µg/mL)	2.68±1.08[a]	1.23±0.64[b]	0.011

*p<0.05 **p<0.001 (Differences in pre and post treatment stages were indicated by lower cases (a and b).

found high (Al-Dawood, 2017). In the study we conducted, we determined that pretreatment SAA concentration in neonatal kids with *Cryptosporidium* spp. was statistically higher than posttreatment (p<0.05). We suggest that the decreased level of post treatment SAA concentration is indicative of the efficacy of the treatment.

In the studies on neonatal lambs, it was reported that Hp concentration increased during the diarrheas due to *Cryptosporidium* spp. (Dinler *et al.,* 2017). Similarly, serum Hp concentrations were high in the calves with diarrhea due to *Cryptosporidium spp.* (Kabu and Uyarlar, 2022; Albayrak and Kabu, 2016). While reviewing the e literature review, we did not find any information regarding the changes in APP values due to *Cryptosporidium spp.* infection in goat kids. However, the increases in the Hp values in *Cryptosporidium spp.* infections in both lambs (Dinler *et al.,* 2017) and calves (Kabu and Uyarlar, 2022; Albayrak and Kabu, 2016) were observed in previous studies which are similar to the results of present study. In our study, Hp concentration was determined to be statistically higher in pretreatment than in posttreatment in neonatal kids with *Crytosporidium spp.* (p<0.05). We are of the opinion that the decrease in the Hp concentration in post treatment stage is indicator of efficacy of the treatment.

CONCLUSION

Acute phase proteins have been used to effectively evaluate the diagnosis, treatment and control mechanisms of various diseases in veterinary as well as in human medicine. Unfortunately, there is only limited information availbale on acute phase response in small ruminants. The present study is of vital importance as it is the first study that provides firsthand information on acute phase response in Cryptosporidiosis affected goat kids.

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Conflict of interest

The authorsdeclare that there is no conflict of interests regarding the publication of this article.

REFERENCES

Albay, M.K., Karakurum, M.C., Sahinduran, S., Sezer, K., Yildiz, R. and Buyukoglu, T. (2014). Selected serum biochemical parameters and acute phase protein levels in a herd of Saanen goats showing signs of pregnancy toxaemia. Veterinarni Medicina. 59(7): 336-342.

Albayrak, H., Kabu, M. (2016). Determining serum haptoglobin and cytokine concentrations in diarrheic calves. Fırat Üniversitesi Sağlık Bilimleri Veteriner Dergisi. 30(2): 113-117.

Al-Dawood, A. (2017). Acute phase proteins as indicators of stress in Baladi goats from Jordan. Acta Agriculturae Scandinavica - Section A: Animal Science. 67(1-2): 58-65.

Cacciò, S.M., Sannella, A.R., Mariano, V., Valentini, S., Berti, F., Tosini, F., Pozio, E. (2013). A rare *Cryptosporidium parvum* genotype associated with infection of lambs and zoonotic transmission in Italy. Veterinary Parasitology. 191(1-2): 128-131.

Ceciliani, F., Ceron, J.J., Eckersall, P.D., Sauerwein, H. (2012). Acute phase proteins in ruminants. Journal of Proteomics. 75(14): 4207-4231.

Cray, C. (2012). Acute phase proteins in animals. Progress in Molecular Biology and Translational Science. 105: 113-150.

Criscitiello, M., Pinsino, A., Field, C.L., Gelain, M.E., Bonsembiante, F. (2019). Acute phase proteins in marine mammals: State of art, perspectives and challenges. Frontiers in Immunology. 10: 1220. https://doi.org/10.3389/fimmu.2019.01220.

Demirbaş, K., Kabu, M. (2021). Evaluation of serum amyloid-A levels in clinically detected pneumonia in goats. Kocatepe Veterinary Journal. 14(4): 430-435.

Dinler, C., Ulutas, B., Voyvoda, H., Ulutas, P.A., Ural, K., Karagenc, T. (2017). Haptoglobin and serum amyloid-A concentrations and their relationship with oocyst count in neonatal lambs experimentally infected with *Cryptosporidium parvum*. Veterinary Parasitology. 247: 49-56.

Feng, Y., Xiao, L. (2017). Molecular Epidemiology of Cryptosporidiosis in China. Frontiers in Microbiology. 8: 1701.

Giadinis, N.D., Papadopoulos, E., Lafi, S.Q., Papanikolopoulou, V., Karanikola, S., Diakou, A., Vergidis, V., Xiao, L., Ioannidou, E., Karatzias, H. (2015). Epidemiological observations on *Cryptosporidiosis* in diarrheic goat kids in Greece. Veterinary Medicine International. 1: 4.

Giadinis, N.D., Symeoudakis, S., Papadopoulos, E., Lafi, S.Q., Karatzias, H. (2012). Comparison of two techniques for diagnosis of cryptosporidiosis in diarrhoeic goat kids and lambs in Cyprus. Tropical Animal Health and Production. 44(7): 1561-1565.

González, F.H.D., Tecles, F., Martínez-Subiela, S., Tvarijonaviciute, A., Soler, L., Cerón, J.J. (2008). Acute phase protein response in goats. Journal of Veterinary Diagnostic Investigation. 20(5): 580-584.

Guo, Y., Li, N., Ryan, U., Feng, Y., Xiao, L. (2021). Small ruminants and zoonotic cryptosporidiosis. Parasitology Research. 120(12): 4189-4198.

Heller, M.C., Johns, J.L. (2015). Acute phase proteins in healthy goats. Journal of Veterinary Diagnostic Investigation. 27(2): 177-181.

Kabu, M., Uyarlar, C. (2022). Determining serum haptoglobin and proinflammatory cytokines concentration in calves clinically diagnosed with pneumonitis, pneumoenteritis and enteritis. Indian J. Anim. Res. 56(4): 498-501.

Kabu, M., Civelek, T., Tunç, A.C., Eser, M. (2023). Evaluating the efficacy of the treatment in lambs with *Cryptosporidium* spp. *via* saa and haptoglobin. Indian J. Anim. Res. doi: 10.18805/IJAR.BF-1582.

Lecchi, C., Dilda, F., Sartorelli, P., Ceciliani, F. (2012). Widespread expression of SAA and Hp RNA in bovine tissues after evaluation of suitable reference genes. Vet. Immunol. Immunopathol. 145(1-2): 556-562.

Ngure, R.M., Eckersall, P.D., Jennings, F.W., Burke, J.M., Stear, M.J., Kennedy, P.G., Murray, M. (1997). Major acute phase response of haptoglobin and serum amyloid-P following experimental infection of mice with *Trypanosoma brucei.* Parasitology International. 46(4): 247-254.

Santin, M. (2020). Cryptosporidium and Giardia in ruminants. Veterinary Clinics of North America: Food Animal Practice. 36(1): 223-238.

Sevgisunar, N.S., Şahinduran, Ş. (2021). Evaluation of some acute phase proteins, cytokines and hepcidin levels in naturally ınfected Saanen goats with paratuberculosis. Mehmet Akif Ersoy Üniversitesi Sağlık Bilimleri Enstitüsü Dergisi. 9(3): 29-37.

Turgay, N. (2011). Özel Boyama Yöntemleri In: Parazitolojide Laboratuvar. 1st edn. [Ok, U.Z., Korkmaz, M. (eds)], Türkiye Parazitoloji Derneği, Türkiye Pp. 23-38.

Wright, S.E., Coop, R.L. (2007). Cryptosporidiosis and Coccidiosis. Diseases of Sheep. 4th edn. Blackwell Publishing, UK. pp. 179-185.